# *In vitro* antioxidant activity and cytotoxicity of sequential extracts from selected black pepper (*Piper nigrum* L.) varieties and *Piper* species

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#### Article history

Received: 16 September 2015 Received in revised form: 9 March 2016 Accepted: 18 March 2016 Abstract

#### **Keywords**

P. nigrum P. longum P. chaba P. colubrinum Antioxidant activity Cytotoxicity Present study evaluated in vitro antioxidant activity and cytotoxicity of four important Piper species (P. nigrum L., P. chaba Hunter, P. longum L. and P. colubrinum Link.) and six black pepper varieties (Sreekara, Subhakara, IISR Malabar Excel, Panniyur-1, Panchami and IISR Thevam). It was performed with sequential extracts of the dried berries/fruits using n-hexane, chloroform, methanol and water in the order of increasing polarity. Concentrated extracts were tested for total phenolic content, in vitro antioxidant activity and cytotoxicity. Methanol and chloroform extracts showed high antioxidant activity than hexane and water extracts. Among black pepper varieties, methanol extract of IISR Malabar Excel followed by that of Panchami and among *Piper* species chloroform extract of *P. colubrinum* expressed highest antioxidant activity. Significant positive correlation between total phenol and antioxidant activity was noted for methanol and chloroform extracts. In vitro cytotoxicity of the extracts was tested on cervical cancer cell line CaSki by MTT assay and compared with that of synthetic anticancer drug Doxorubicin. Results showed more cytotoxicity with more extract and increased time of exposure with CaSki. Chloroform extract of P. longum and P. colubrinum were found to be highly toxic to CaSki than other extracts. By considering three time intervals, chloroform extract of IISR Malabar Excel was more toxic to CaSki than other black pepper varieties. To the best of our knowledge, this is the first report regarding in vitro antioxidant activity and cytotoxicity on CaSki for sequential extracts of *P. colubrinum* fruits cultivated in India. This is also the first report on variability in antioxidant activity and cytotoxicity (on CaSki) of sequential extracts from black pepper varieties selected for the study.

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### Introduction

Human body produces free radicals mainly Reactive Oxygen Species (ROS) as a part of normal metabolic processes. Mitochondria, peroxisomes and immune cells like leukocytes and macrophages are the main endogenous sources for free radical production in cells. Many acute and chronic diseases like cancer, diabetes, inflammation, arthritis, aging, atherosclerosis and various neurodegenerative disorders mainly arises from oxidative stress initiated by highly reactive and unstable free radicals. So, the oxidation and antioxidation balance should be maintained for a healthy biological system. This can be achieved by exploring compounds with antioxidant activity (Agbor et al., 2006; Krishnaswami et al., 2013). Plants are tremendous source for such antioxidants. Several studies revealed that various phytochemicals especially phenolic compounds possess remarkable antioxidant activity (Van Acker et al., 1996; Pietta, 2000; Williams et al., 2004; Chatterjee et al., 2007). Thus, in recent

years, researchers mainly focused on isolation and identification of such antioxidants from various plant species and looking for new leads to develop better drugs from these phytochemicals for various diseases by oxidative stress. It may help to reduce the risk of using synthetic antioxidant and anticancer drugs. *Piper* is one of such plant genera with diverse medicinal properties. *P. nigrum, P. longum, P. chaba* and *P. colubrinum* are the *Piper* species selected for the study.

*P. nigrum* (Black pepper) is an important spice valued for its aroma and pungency. Aroma is contributed by essential oil constituents and pungency by the alkaloid piperine. Black pepper and its alkaloid piperine is known for its therapeutic properties like antimicrobial, analgesic, antipyretic, antioxidant, anticancer and also for enhancement of bioavailability of drugs (Lee *et al.*, 1984; Bano *et al.*, 1991; Vijayakumar *et al.*, 2004; Karsha and Lakshmi, 2010; Pingili *et al.*, 2012). *P. longum* (Long pepper) and *P. chaba* has been used in traditional medicine and are the major ingredients in many Ayurvedic



systems. Their antimicrobial activity, antioxidant activity, antitumor effect and efficacy against respiratory tract disorders, rheumatic pains and diarrhoea were studied (Anshuman *et al.*, 1984; Yusuf *et al.*, 1994; Srinivasa *et al.*, 2001; Sunila and Kuttan, 2004; Taufiq-Ur-Rahman *et al.*, 2005; Samudram *et al.*, 2009; Rahman *et al.*, 2011). *P. colubrinum* is an exotic *Piper* species native to Northern part of South America. It has great importance because of its resistance to *Phytophthora capsici* and *Radopholus similis* (Ravindran and Remashree, 1998). But it is to be noted that medicinal values of *P. colubrinum* Link. is under explored.

Present study is aimed to find *in vitro* antioxidant activity and cytotoxicity of sequential extracts from four *Piper* species *viz*. *P. nigrum*, *P. longum*, *P. chaba* and *P. colubrinum* and to find out correlation between total phenolics of different extracts of this species with different antioxidant assays. *In vitro* cytotoxicity of sequential extracts was checked on cervical cancer cell line CaSki since it is one of the least studied cancer cell line among these *Piper* species.

#### **Materials and Methods**

#### Collection of samples

Matured and dried fruits/berries of *P. nigrum* (wild), *P. longum, P. chaba, P. colubrinum* and six high yielding black pepper varieties (Sreekara, Subhakara, IISR Malabar Excel, IISR Thevam, Panchami and Panniyur-1) were collected from ICAR-Indian Institute of Spices Research (IISR) Experimental Farm, Peruvannamuzhi, Kerala, India and used for the study.

#### Chemicals

Folin-Ciocalteau reagent, ammonium molybdate, ascorbic acid, potassium ferricyanide, trichloroacetic acid (TCA), ferric chloride, Ethylene diamine ferrozine. tetraaceticacid (EDTA) and dimethylsulphoxide (DMSO) were purchased from Sisco Research Laboratories (Mumbai, India). Gallic acid, 1, 1-diphenyl-2picryl-hydrazyl (DPPH), butylated hydroxyanisole (BHA) and 3-(4,5--dimethylthiazole-2-yl)-2,5diphenyltetrazolium bromide (MTT) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Fetal bovine serum (FBS), Dulbecco's modified Eagle's medium (DMEM) and antibiotic antimycotic mixture were purchased from Life Technologies (Carlsbad, CA, USA). All other reagents and chemicals used were of analytical grade and of highest purity.

### Preparation of extracts

All the samples were powdered and extracted sequentially with n-hexane, chloroform, methanol and water in the increasing order of polarity using Soxhlet apparatus. After the completion of extraction with one solvent, the sample left was dried at room temperature and extracted with next solvent. Each extract was filtered and evaporated to dryness using a BUCHI rotary evaporator equipped with BUCHI rotavapor R-205 and BUCHI heating bath B-490. The concentrated extracts were stored at 4°C until analysis. Sequential extraction helps for effective and complete extraction of compounds with different polarity (Policegoudra *et al.*, 2011).

### Determination of total phenol content

Total phenolic content of all extracts was estimated by Folin-Ciocalteau method (Malick and Singh, 1980). In this method, phenolics reacts with phosphomolybdic acid in Folin-Ciocalteau reagent in alkaline medium and produce blue coloured molybdenum blue which is measured at 650 nm and the results were expressed as milligram gallic acid equivalents/g of extract (mg GAE/g of extract).

#### In vitro antioxidant activity

Antioxidant activity was tested using 1,1-diphenyl-2-picryl hydrazyl (DPPH) free radical scavenging activity, Phosphomolybdenum method, Ferric reducing power method and Ferrous chelating activity. Stock solution of all the extracts and synthetic antioxidant Butylated Hydroxyanisole (BHA) was prepared in methanol with a concentration of 10 mg/ mL.

### *Free radical scavenging activity on 1,1-diphenyl-2picryl hydrazyl (DPPH)*

The antioxidant activity of the extracts was determined in terms of their hydrogen donating or radical scavenging ability and thus to scavenge DPPH free radical (Braca et al., 2001). A working solution with a concentration range of 10 - 500  $\mu$ g/ mL for methanol and chloroform extracts and 100 - 2000  $\mu$ g/mL for hexane and water extracts were prepared from the respective stock solutions. One mL of each aliquot was added into test tubes and final volume was made up to 4 mL with methanol. One mL of 0.004% DPPH was added to the samples. After proper mixing, samples were incubated at dark for 30 minutes and absorbance was taken at 517 nm with UV-VIS spectrophotometer (UV-1800, Shimadzu Corp., Japan). The synthetic antioxidant BHA was taken as positive control. Radical scavenging ability of each extract was expressed as  $IC_{50}$  value, which represents the amount of extract required to scavenge

50% DPPH free radicals. IC  $_{\rm 50}$  value was calculated using statistical software SAS 9.3 and expressed as  $\mu g/mL.$ 

# Total antioxidant activity by Phosphomolybdenum method

An aliquot of 50 µL for methanol and chloroform extracts and 100 µL for hexane and water extracts were taken from the corresponding stock solutions and the final volume was made up to 3 mL with methanol. One mL of phosphomolybdenum reagent (0.6 M sulphuric acid, 28 mM disodium hydrogen phosphate, 4 mM ammonium molybdate) was added into the tubes and incubated at 95°C for 90 minutes. After incubation, the samples were read at 695 nm using UV-VIS spectrophotometer (UV-1800, Shimadzu Corp., Japan). Ascorbic acid (0.2-1.0 mM) was used as standard for the preparation of calibration curve. The results were expressed as molar ascorbic acid equivalents/g of extract (M AAE/ g of extract). Synthetic antioxidant BHA was used for comparison (Prieto et al., 1999).

#### Ferric reducing power (FRP) method

The reductive capacity of extracts was estimated by the method described by Oyaizu (1986). 50 µL of methanol, chloroform and hexane extracts and 100 µL of water extract were taken from the corresponding stock solutions and the final volume was made up to 1 mL with distilled water. The samples were then mixed with phosphate buffer (0.2 M, pH 6.6). Potassium ferricyanide (2.5 mL, 1% W/V) was added to the mixture and incubated at 50°C for 30 min. The reaction was terminated by adding trichloroacetic acid (10% W/V) and the mixture was centrifuged at 3000 rpm for 20 minutes. The supernatant was mixed with distilled water and FeCl<sub>2</sub> (0.1% W/V) solution and the absorbance was measured at 700 nm using UV-VIS spectrophotometer (UV-1800, Shimadzu Corp., Japan). Ascorbic acid (0.25-1.0 mM) was used as standard for the preparation of calibration curve. Increased absorbance of the reaction mixture indicated greater reducing power and it was expressed in molar ascorbic acid equivalents/g of extract (M AAE/ g of extract). Synthetic antioxidant BHA was kept as positive control.

#### Determination of ferrous chelating ability

The ability of extracts to chelate ferrous ion was estimated by measuring the intensity of ferrous-ferrozine complex (Carter, 1971). An aliquot of 25  $\mu$ L for methanol and chloroform extracts and 50  $\mu$ L for hexane and water extracts from the stock solutions were taken into test tubes and made up the volume to

3 mL with methanol. All the test solutions were then treated with  $\text{FeCl}_2$  (0.1 mL; 2 mM). After incubation for 5 minutes, 0.4 mL of 5 mM ferrozine was added to the above mixture. After incubation for 10 minutes at room temperature, the absorbance was recorded at 562 nm with UV-VIS spectrophotometer (UV-1800, Shimadzu Corp., Japan). EDTA (10-50 µg) was used as standard for the preparation of calibration curve. Ferrous chelating ability was expressed as milligram EDTA/g of extract (mg EDTA/g of extract).

#### In vitro cytotoxicity analysis

In vitro cytotoxicity study of sequential extracts was tested on cervical cancer cell line CaSki by MTT assay. Cervical cancer cell line CaSki was cultured as adherent monolayer as per earlier method (Freshney, 2007) and maintained in 90% Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and incubated at 37°C and 5% CO<sub>2</sub>. Stock solution (250 mg/mL) of each extract was prepared in DMSO and stored at -20°C until use.

#### MTT Assay

MTT (3-(4,5))Dimethylthiazol-2yl)-2,5-Diphenyl Tetrazolium Bromide) assay (Mosmann, 1983) is based on the ability of a mitochondrial dehydrogenase enzyme of viable cells to cleave the tetrazolium rings of the pale yellow tetrazole MTT to a dark blue formazan crystals. Formazan crystals are impermeable to cell membranes which results in its accumulation within healthy cells. The crystals can be solubilised by adding lysis buffer and its colour can be measured spectrophotometrically. The level of the coloured formazan product is directly proportional to the number of surviving cells. The stock solutions of extracts were diluted in 10% DMEM to get lower concentration of extracts (0.5 mg/mL). Cells harvested in the log phase of growth were counted and seeded ( $5 \times 10^3$  cells/well) in 96 well micro titer plates and incubated over night at 37°C in a humidified 5% CO<sub>2</sub> incubator (NuAire, USA). The cells were then allowed to react with different amount of extracts (25, 50 and 100 µg) for 24, 48 and 72 hrs in a humidified 5% CO<sub>2</sub> incubator (NuAire, USA) at 37°C. Synthetic anticancer drug Doxorubicin served as positive control whereas 10% DMEM was taken as negative control. After incubation, the medium was discarded and wells were washed with PBS. 100 µL of the MTT (5% in 10% DMEM media) was added and incubated for 2 hours. MTT lysis buffer (100  $\mu$ L) was added to solubilise the colored formazan crystals formed by the reduction of MTT. After incubated for 4 hrs, the absorbance was measured at 570 nm using a micro plate spectrophotometer (Bio-Tek, USA).

The percentage cytotoxicity was determined and IC<sub>50</sub> (amount of extracts required for 50% cytotoxicity) values were calculated from the dose response curve for CaSki. Potential extracts were again put for MTT assay and their IC<sub>50</sub> was calculated by using Microsoft Excel 2007 and compared with synthetic anticancer drug Doxorubicin.

# Preliminary phytochemical analysis of screened extracts

The extracts screened for high antioxidant activity and *in vitro* cytotoxicity were subjected to preliminary phytochemical screening. The extracts were tested for phenolics, alkaloids, flavones, flavonols, steroids/triterpenes, saponins, fixed oils and fats, carbohydrates and protein by adopting standard protocols (Trease and Evans, 2002; Khandelwal, 2008; Kokate *et al.*, 2008).

### Statistical analysis

Data were combined and analyzed by analysis of variance (ANOVA). The ANOVA was performed with the MSTATC software (version 1.41). The Significant differences (p<0.05) was estimated by Duncan's multiple range test (DMRT) using 'RANGE' procedure and the correlation coefficient (r) was determined using the 'CORR' procedure of MSTATC. Values were expressed as Mean  $\pm$  SD of three replicates and superscripts represent significant difference as per DMRT (p < 0.05).

#### **Results and Discussion**

#### Total phenolic content of extracts

It is reported that phenolics especially phenolic acids and flavonoids shows antioxidant activity and they are responsible for variation in antioxidant activity of plants (Luo et al., 2004; Demiray et al., 2009). Phenolics exhibit antioxidant activity by various mechanisms like radical scavenging activity, transition metal chelating activity and lipid peroxidation inhibition. Due to this multiple mechanism of antioxidant activity, phenolics became an interesting class of compounds for researchers to find natural health beneficial phytochemicals (Rice-Evans et al., 1997; Chen and Ahn, 1998; Yanishlieva and Marinova, 2001). Thus, estimation of total phenol from the selected extracts is crucial. Table 1 showed variability in total phenol content among four extracts from different black pepper varieties and Piper species. Chloroform and methanol extracts showed maximum phenol content than hexane and water extracts. Hexane extract of P. colubrinum also showed high phenolic content. Total phenol

content was in the range of  $3.14 \pm 0.13$  to  $30.7\pm1.23$ ,  $14.02\pm0.56$  to  $100.6\pm4.02$ ,  $14.86\pm0.59$  to  $50.85\pm2.03$  and  $1.05 \pm0.04$  to  $5.12\pm0.20$  mg GAE/g of extract in hexane, chloroform, methanol and water extracts respectively. Chloroform extract of *P. colubrinum* followed by methanol extract of IISR Malabar Excel showed highest total phenol among all the extracts. Among black pepper varieties, methanol extract of IISR Malabar Excel showed highest total phenol extract of Panchami. It was clear from the table 1 that the extracts showed significant variability in their total phenol content among black pepper varieties and also among *Piper* species.

#### DPPH free radical scavenging activity

Table 2 showed the inhibitory concentration of each extract required to scavenge 50% DPPH free radicals (IC<sub>50</sub> in  $\mu$ g/mL). Though all the extracts exhibited free radical scavenging ability, chloroform and methanol extracts predominated. Hexane extract of P. colubrinum also showed high DPPH radical scavenging ability. IC<sub>50</sub> value for chloroform and methanol extracts was found to be very low (27.4  $\pm 1.1$  to 228.9 $\pm 9.2$  µg/mL for chloroform extract and  $29.42\pm1.2$  to  $153.9\pm6.2$  µg/mL for methanol extract) and this indicated their high ability to scavenge DPPH free radicals. Among methanol extracts, IISR Malabar Excel and among chloroform extracts, P. colubrinum showed highest radical scavenging activity. Methanol extract of Panchami and P. colubrinum also showed high ability to scavenge DPPH. Hexane extracts (except P. colubrinum) and water extracts showed comparatively low DPPH radical scavenging ability with high IC<sub>50</sub> values. Significant variation was observed among varieties and also among Piper species in DPPH radical scavenging ability. DPPH radical scavenging activity of BHA was also checked and compared with that of extracts. It was found that BHA was superior to extracts for scavenging DPPH free radicals (IC<sub>50</sub> - 5.2 $\mu g/mL$ ).

DPPH free radical scavenging ability of samples is due to their hydrogen donating ability. DPPH inhibitory capacity of water and methanol extracts of black pepper was reported as 35.2 and 45.66% (Gupta *et al.*, 2014). Ethyl acetate and water extract of black pepper showed a concentration dependent increase in their DPPH radical scavenging ability (Asimi *et al.*, 2013). Methanolic extract of black pepper showed good DPPH scavenging ability with  $IC_{50}$  value of 144.1±2.2 µg/mL (Nooman *et al.*, 2008). DPPH scavenging ability of ethanol extract of black pepper from Brazil was reported with EC<sub>50</sub>

Sample Hexane Chloroform Methanol Water Sreekara 5.98±0.24° 22.32±0.89° 17.44±0.70de 3.41±0.14° Subhakara 4.90±0.20de 14.02±0.56e 21.28±0.85° 1.73±0.07° **IISR Malabar Excel** 7.74±0.31<sup>b</sup> 35.26±1.41<sup>b</sup> 50.85±2.03ª 1.12±0.04<sup>f</sup> 4.26±0.17<sup>e</sup> 21.14±0.85° 20.72±0.83° IISR Theyam 5.12±0.20ª 6.98±0.28<sup>b</sup> 19.33±0.77<sup>cd</sup> Panniyur-1 17.26±0.69d 1.62±0.06<sup>e</sup> 7.43±0.30b 38.74±1.55<sup>b</sup> Panchami 23.64±0.95° 1.12±0.04<sup>f</sup> 3.14±0.13<sup>f</sup> 15.06±0.60de 14.86±0.59<sup>f</sup> 3.84±0.15<sup>b</sup> P. nigrum (wild) P. longum 5.41±0.22<sup>∞</sup> 15.66±0.63de 17.38±0.70de 2.16±0.09<sup>d</sup> 5.37±0.21<sup>∞</sup> 16.50±0.66de 16.53±0.66ef 2.22±0.09d P. chaba P. colubrinum 30.7±1.23ª 100.6±4.02ª 37.26±1.49<sup>b</sup> 1.05±0.04<sup>f</sup>

Table 1. Total Phenolic content from different extracts (mg GAE/g of extract)

value 110±2 g spice/kg DPPH (Mariutti et al., 2008). Petroleum ether extract of black pepper was also subjected to DPPH scavenging activity and found that there is a concentration dependent increase in their scavenging activity and hence black pepper could be considered as a potent source of natural antioxidant (Singh et al., 2008). IC<sub>50</sub> for DPPH radical scavenging activity of chloroform, ethyl acetate, hexane, ethanol, hydroethanol and aqueous extracts of *P. longum* were reported as 6,54, 70, 50, 26, 19.5 µg/mL respectively (Barua et al., 2014). A strong bioactive alkaloid Chabbarin was isolated from acetone extract of *P. chaba* and its strong ability to scavenge DPPH was reported with a very less  $IC_{50}$ value of 3 µg/mL (Biswas et al., 2012). Antioxidant activity of methanolic extract of P. longum and P. chaba was estimated by various assays including DPPH radical scavenging activity and showed that P. longum is superior to P. chaba (Ramesh kumar et al., 2011).

# Total antioxidant activity by Phosphomolybdenum method

Table 2 showed total antioxidant capacity of each extract by Phosphomolybdenum method. The assay is based on the ability of sample to reduce Mo (VI) to Mo (V) and the resultant green colour was measured. Hexane extract was in the range of 0.41±0.08 to 0.75±0.09 M AAE/g of extract whereas chloroform, methanol and water extracts were in the range of 0.81±0.26 to 1.85±0.01, 0.65±0.13 to 1.87±0.01 and 0.09±0.01 to 0.49±0.13 M AAE/ g of extract. All methanol and chloroform extracts and also the hexane extract of *P. colubrinum* showed high total antioxidant activity whereas hexane extract of other samples and all water extracts expressed comparatively low activity. Methanol extract of IISR Malabar Excel and chloroform extract of P. colubrinum showed highest antioxidant activity. The result of extracts was compared with that of synthetic standard BHA and found that BHA showed high ability to reduce Molybdenum (4.6 Molar AAE/ g of extract). The ability of cold methanolic extract from P. nigrum and P. longum to reduce molybdenum was

reported earlier (Prasad and Sushant, 2014). *Ferric reducing power (FRP) method* 

Antioxidant activity was also checked on the basis of the ability of antioxidants to reduce ferric (III) ion to ferrous (II) ion. Table 3 showed ferric reducing capacity of each extract. Among hexane extracts, P. colubrinum showed highest activity followed by IISR Malabar Excel and Panchami. Chloroform and methanol extracts were in the range of  $0.54\pm0.01$ to 1.02±0.16 and 0.50±0.01 to 1.18±0.02 M AAE/ g of extract respectively. For chloroform extract, P. colubrinum followed by IISR Malabar Excel and for methanol extract, IISR Malabar Excel followed by Panchami showed highest ferric reducing power. Water extract showed comparatively very low activity. Among all extracts, chloroform extract of P. colubrinum and methanol extract of IISR Malabar Excel showed highest ferric reducing power ability. Ferric reducing ability of different extracts from above selected *Piper* species were also reported by researchers (Kapoor et al., 2009; Gopalakrishna et al., 2010).

#### Ferrous chelating activity

The ability to chelate ferrous ion was in the range of 72.17±2.9 to 137±5.5, 228.5±9.14 to 320.1±12.8, 245.5±9.82 to 349.8±13.9 and 124.5±4.9 to 170.6±6.8 mg EDTA/g of extract for hexane, chloroform, methanol and water respectively (Table 3). Hexane and water extracts showed comparatively low ferrous chelating ability whereas it was found to be high for methanol and chloroform extracts. Methanol extract of IISR Malabar Excel was found to be superior for their ability to chelate ferrous ions. Chloroform extract of *P. colubrinum*, methanol extract of Panchami and methanol extract of *P. colubrinum* were also showed high ferrous chelating ability.

Secondary metabolites like phenolics and flavonoids can chelate metal ions and often decrease the prooxidant activity of metal ions. Metal chelating potency of phenolic compounds is dependent upon their unique phenolic structure and the number and location of the hydroxyl groups (Van Acker *et al.*, 1998; Santoso *et al.*, 2004). Metal chelating ability

DPPH scavenging activity of sequential extracts in terms of IC <sub>50</sub> (µg/mL)					
Sample	Hexane	Chloroform Methanol		Water	
Sreekara	1068.0±42.7 <sup>d</sup>	148.2±5.9 <sup>e</sup>	62.45±2.5°	935.47±37.4 <sup>e</sup>	
Subhakara	1240.0±49.6 <sup>c</sup>	228.9±9.2ª	54.09±2.2 <sup>d</sup>	800.00±32.0 <sup>f</sup>	
IISR Malabar Excel	776.54±31.1 <sup>f</sup>	99.61±3.9 <sup>f</sup>	29.42±1.2 <sup>f</sup>	1942.0±77.7ª	
IISR Thevam	1430.0±57.2 <sup>b</sup>	154.2±6.2 <sup>de</sup>	54.04±2.2 <sup>d</sup>	765.20±30.6 <sup>f</sup>	
Panniyur-1	1019.0±40.8 <sup>de</sup>	170.1±6.8 <sup>bc</sup>	54.58±2.2 <sup>d</sup>	1784.0±71.4 <sup>b</sup>	
Panchami	970.00±38.8 <sup>e</sup>	104.6±4.2 <sup>f</sup>	40.14±1.6 <sup>e</sup>	1620.0±64.8 <sup>c</sup>	
P. nigrum (wild)	1830.0±73.2ª	164.9±6.6 <sup>bcd</sup>	153.9±6.2ª	1025.0±41.0 <sup>e</sup>	
P. longum	1400.0±56.0 <sup>b</sup>	174.1±6.9 <sup>b</sup>	130.4±5.2 <sup>b</sup>	1660.0±66.4 <sup>c</sup>	
P. chaba	1800.0±72.0ª	159.9±6.4 <sup>cd</sup>	152.1±6.1ª	1900.0±76.0 <sup>a</sup>	
P. colubrinum	122.70±4.91 <sup>g</sup>	27.40±1.1 <sup>g</sup>	42.28±1.7 <sup>e</sup>	1207.0±48.3 <sup>d</sup>	
BHA	5.200±0.21				
Total antioxidant capacity by Phosphomolybdenum method (M AAE/ g of extract)					
Total antioxidant capacit	y by Phosphomoly	ybdenum metho	d (MAAE/gof	extract)	
Total antioxidant capacit Sample	y by Phosphomoly Hexane	ybdenum metho Chloroform	d (M AAE/g of Methanol	extract) Water	
Total antioxidant capacit Sample Sreekara	y by Phosphomoly Hexane 0.59±0.01 <sup>bc</sup>	ybdenum metho Chloroform 1.19±0.19 <sup>bc</sup>	d (M AAE/ g of <u>Methanol</u> 0.83±0.23 <sup>d</sup>	extract) Water 0.32±0.01 <sup>bc</sup>	
Total antioxidant capacit Sample Sreekara Subhakara	y by Phosphomoly <u>Hexane</u> 0.59±0.01 <sup>bc</sup> 0.53±0.06 <sup>bcd</sup>	ybdenum metho Chloroform 1.19±0.19 <sup>bc</sup> 1.08±0.04 <sup>bc</sup>	d (M AAE/ g of <u>Methanol</u> 0.83±0.23 <sup>d</sup> 0.85±0.03 <sup>d</sup>	extract) Water 0.32±0.01 <sup>bc</sup> 0.24±0.07 <sup>cd</sup>	
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Total antioxidant capacit Sample Sreekara Subhakara IISR Malabar Excel IISR Thevam Panniyur-1 Panchami <i>P. nigrum</i> (wild)	y by Phosphomoly Hexane 0.59±0.01 <sup>bc</sup> 0.53±0.06 <sup>bcd</sup> 0.62±0.09 <sup>b</sup> 0.51±0.04 <sup>bcd</sup> 0.55±0.01 <sup>bcd</sup> 0.64±0.01 <sup>ab</sup> 0.41±0.08 <sup>d</sup>	ybdenum metho Chloroform 1.19±0.19 <sup>bc</sup> 1.08±0.04 <sup>bc</sup> 1.06±0.07 <sup>bc</sup> 0.81±0.26 <sup>c</sup> 0.83±0.02 <sup>bc</sup> 1.24±0.12 <sup>b</sup> 1.01±0.25 <sup>bc</sup>	d (M AAE/ g of <u>Methanol</u> 0.83±0.23 <sup>d</sup> 0.85±0.03 <sup>d</sup> 1.87±0.01 <sup>a</sup> 1.13±0.04 <sup>c</sup> 0.66±0.10 <sup>d</sup> 1.52±0.21 <sup>b</sup> 0.73±0.08 <sup>d</sup>	extract) Water 0.32±0.01 <sup>bc</sup> 0.24±0.07 <sup>cd</sup> 0.09±0.01 <sup>e</sup> 0.49±0.13 <sup>a</sup> 0.24±0.01 <sup>cd</sup> 0.15±0.01 <sup>de</sup> 0.45±0.06 <sup>ab</sup>	
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Total antioxidant capacit Sample Sreekara Subhakara IISR Malabar Excel IISR Thevam Panniyur-1 Panchami <i>P. nigrum</i> (wild) <i>P. longum</i> <i>P. chaba</i>	y by Phosphomoly Hexane $0.59\pm0.01^{bc}$ $0.53\pm0.06^{bcd}$ $0.62\pm0.09^{b}$ $0.51\pm0.04^{bcd}$ $0.55\pm0.01^{bcd}$ $0.64\pm0.01^{ab}$ $0.41\pm0.08^{d}$ $0.43\pm0.06^{d}$ $0.47\pm0.01^{cd}$	ybdenum metho <u>Chloroform</u> 1.19±0.19 <sup>bc</sup> 1.08±0.04 <sup>bc</sup> 1.06±0.07 <sup>bc</sup> 0.81±0.26 <sup>c</sup> 0.83±0.02 <sup>bc</sup> 1.24±0.12 <sup>b</sup> 1.01±0.25 <sup>bc</sup> 1.16±0.34 <sup>bc</sup> 0.85±0.06 <sup>bc</sup>	d (M AAE/ g of <u>Methanol</u> 0.83±0.23 <sup>d</sup> 0.85±0.03 <sup>d</sup> 1.87±0.01 <sup>a</sup> 1.13±0.04 <sup>c</sup> 0.66±0.10 <sup>d</sup> 1.52±0.21 <sup>b</sup> 0.73±0.08 <sup>d</sup> 0.65±0.13 <sup>d</sup> 0.79±0.08 <sup>d</sup>	extract) Water 0.32±0.01 <sup>bc</sup> 0.24±0.07 <sup>cd</sup> 0.09±0.01 <sup>e</sup> 0.49±0.13 <sup>a</sup> 0.24±0.01 <sup>cd</sup> 0.15±0.01 <sup>de</sup> 0.45±0.06 <sup>ab</sup> 0.19±0.03 <sup>de</sup> 0.21±0.01 <sup>cde</sup>	
Total antioxidant capacit Sample Sreekara Subhakara IISR Malabar Excel IISR Thevam Panniyur-1 Panchami <i>P. nigrum</i> (wild) <i>P. longum</i> <i>P. chaba</i> <i>P. colubrinum</i>	y by Phosphomoly Hexane $0.59\pm0.01^{bc}$ $0.53\pm0.06^{bcd}$ $0.62\pm0.09^{b}$ $0.51\pm0.04^{bcd}$ $0.55\pm0.01^{bcd}$ $0.64\pm0.01^{ab}$ $0.41\pm0.08^{d}$ $0.43\pm0.06^{d}$ $0.47\pm0.01^{cd}$ $0.75\pm0.09^{a}$	ybdenum metho <u>Chloroform</u> 1.19±0.19 <sup>bc</sup> 1.08±0.04 <sup>bc</sup> 1.06±0.07 <sup>bc</sup> 0.81±0.26 <sup>c</sup> 0.83±0.02 <sup>bc</sup> 1.24±0.12 <sup>b</sup> 1.01±0.25 <sup>bc</sup> 1.16±0.34 <sup>bc</sup> 0.85±0.06 <sup>bc</sup> 1.85±0.01 <sup>a</sup>	d (M AAE/ g of <u>Methanol</u> 0.83±0.23 <sup>d</sup> 0.85±0.03 <sup>d</sup> 1.87±0.01 <sup>a</sup> 1.13±0.04 <sup>c</sup> 0.66±0.10 <sup>d</sup> 1.52±0.21 <sup>b</sup> 0.73±0.08 <sup>d</sup> 0.65±0.13 <sup>d</sup> 0.79±0.08 <sup>d</sup> 1.15±0.03 <sup>c</sup>	extract) Water $0.32\pm0.01^{bc}$ $0.24\pm0.07^{cd}$ $0.09\pm0.01^{e}$ $0.49\pm0.13^{a}$ $0.24\pm0.01^{cd}$ $0.15\pm0.01^{de}$ $0.45\pm0.06^{ab}$ $0.19\pm0.03^{de}$ $0.21\pm0.01^{cde}$ $0.33\pm0.01^{bc}$	

Table 2. DPPH radical scavenging activity and Total antioxidant capacity of sequential extracts

of water and ethanolic extract of black pepper was reported as 84±2.20% and 83±4.36% respectively (Gulcin, 2005).

# Correlation between total phenolic content and antioxidant activity

Several studies reported that phenolic constituents in spices and other plants have significant antioxidant properties (Shan et al., 2005; Wu et al., 2006; Maizura et al., 2011). In the present study, chloroform and methanol extracts were screened for high antioxidant activity by different assays. So correlation of total phenolic content of these two extracts from all the samples with their antioxidant activity by each assay was performed. Result showed significant (p < 0.05) negative correlation between total phenolic content of each extract and their  $IC_{50}$  value to scavenge DPPH free radicals. Correlation coefficient (r) for total phenolic content of chloroform extract and their  $IC_{50}$ for DPPH scavenging activity was -0.86 and that for total phenolic content of methanol extract and their IC<sub>50</sub> for DPPH scavenging activity was -0.72. This indicated that, extract with high total phenol shows less IC<sub>50</sub> and thus more ability to scavenge DPPH free radicals. Correlation coefficient (r) for total phenolic content of chloroform extract with their antioxidant activity by phosphomolybdenum method, FRP and ferrous chelating activity was +0.86, +0.81 and +0.95

respectively (p<0.05). Total phenolic content of methanol extracts of samples also showed significant (p<0.05) positive correlation with their antioxidant activity by phosphomolybdenum method (r=+0.94), FRP (r=+0.92) and ferrous chelating activity (r=+0.79). High ability of *P. colubrinum* hexane extract to scavenge DPPH may also be an indication of radical scavenging ability of phenolics since that extract contains more total phenol than other hexane extracts. Such linearity between total phenol of black pepper extracts and their antioxidant activity was reported by researchers (Gulcin, 2005; Nahak and Sahu, 2011).

# In vitro *cytotoxicity of sequential extracts from* Piper *species and black pepper varieties.*

To screen the extracts with high cytotoxicity to CaSki, MTT assay was performed using all extracts with a mass range of 25-100  $\mu$ g for three time intervals viz. 24, 48 and 72 hrs. All the extracts showed cytotoxicity (%) in a dose dependent and time dependent manner. Among all extracts tested, chloroform extracts of all samples and hexane extract of *P. colubrinum* expressed more cytotoxicity. IC<sub>50</sub> value was calculated for chloroform extracts of black pepper and wild *P. nigrum* and they have categorised by Duncan's multiple range test (DMRT) to screen black pepper chloroform extract with

FRP Method (M AAE/ g of extract)					
Sample	Hexane	Chloroform	Methanol	Water	
Sreekara	0.54±0.04 <sup>cde</sup>	0.64±0.04 <sup>cde</sup>	0.61±0.01 <sup>f</sup>	0.27±0.02 <sup>ab</sup>	
Subhakara	0.57±0.04 <sup>bcd</sup>	0.54±0.01 <sup>e</sup>	0.75±0.01 <sup>de</sup>	0.23±0.01 <sup>cd</sup>	
IISR Malabar Excel	0.63±0.05 <sup>ab</sup>	0.91±0.07 <sup>ab</sup>	1.18±0.02 <sup>a</sup>	0.22±0.01 <sup>de</sup>	
IISR Thevam	0.59±0.01 <sup>abc</sup>	0.78±0.04 <sup>bc</sup>	0.81±0.05 <sup>cd</sup>	0.29±0.01 <sup>a</sup>	
Panniyur-1	0.59±0.04 <sup>abc</sup>	0.73±0.07 <sup>cd</sup>	0.71±0.04 <sup>e</sup>	0.25±0.01 <sup>bc</sup>	
Panchami	0.63±0.05 <sup>ab</sup>	0.76±0.01 <sup>bc</sup>	0.93±0.06 <sup>b</sup>	0.21±0.01 <sup>e</sup>	
P. nigrum (wild)	0.46±0.01 <sup>e</sup>	0.55±0.12 <sup>e</sup>	0.52±0.02 <sup>g</sup>	0.26±0.01 <sup>b</sup>	
P. long um	0.49±0.03 <sup>e</sup>	0.59±0.04 <sup>de</sup>	0.50±0.01 <sup>g</sup>	0.22±0.01 <sup>de</sup>	
P. chaba	0.52±0.04 <sup>de</sup>	0.57±0.03 <sup>de</sup>	0.56±0.01 <sup>fg</sup>	0.22±0.01 <sup>de</sup>	
P. colubrinum	0.65±0.04 <sup>a</sup>	1.02±0.16 <sup>a</sup>	0.84±0.05 <sup>c</sup>	0.21±0.01 <sup>e</sup>	
BHA	3.46±0.14				
Ferrous chelating activ	ity (mg EDTA/g	ofextracts)			
Sample	Hexane	Chloroform	Methanol	Water	
Sreekara	130.5±5.22 <sup>b</sup>	232.2±9.29 <sup>cd</sup>	294.5±11.8 <sup>cde</sup>	167.7±6.7 <sup>ab</sup>	
Subhakara	108.6±4.3 <sup>d</sup>	228.5±9.14 <sup>d</sup>	314.7±12.6 <sup>bc</sup>	157.8±6.3 <sup>bc</sup>	
IISR Malabar Excel	131.7±5.3 <sup>ab</sup>	251.6±10.1 <sup>b</sup>	349.6±13.9 <sup>a</sup>	126.6±5.1 <sup>e</sup>	
IISR Thevam	131.1±5.2 <sup>♭</sup>	234.3±9.37 <sup>bcd</sup>	297.2±11.9 <sup>cd</sup>	170.6±6.8ª	
Panniyur-1	133.6±5.3 <sup>ab</sup>	230.3±9.21 <sup>∞</sup>	316.5±12.7 <sup>bc</sup>	153.4±6.1 <sup>cd</sup>	
Panchami	132.8±5.3 <sup>ab</sup>	249.3±9.97 <sup>bc</sup>	320.8±12.8 <sup>b</sup>	143.4±5.7 <sup>d</sup>	
P. nigrum (wild)	72.19±2.9 <sup>e</sup>	241.6±9.66 <sup>bcd</sup>	245.5±9.82 <sup>f</sup>	166.2±6.7 <sup>ab</sup>	
P. long um	111.4±4.5 <sup>d</sup>	249.1±9.96 <sup>bc</sup>	273.4±10.9 <sup>e</sup>	147.9±5.9 <sup>cd</sup>	
P. chaba	123.5±4.9°	243.3±9.73 <sup>bcd</sup>	290.8±11.6 <sup>de</sup>	153.2±6.1 <sup>cd</sup>	
P. colubrinum	137.0±5.5ª	320.1±12.8ª	319.8±12.8 <sup>b</sup>	124.5±4.9 <sup>e</sup>	
BHA					

Table 3. Ferric reducing power (FRP) and ferrous chelating activity of sequential extracts

highest cytotoxicity. Result showed that chloroform extract of IISR Malabar Excel showed lowest IC<sub>50</sub> value and thus highest cytotoxicity to CaSki for all the three time intervals (Table 4 A). Thus chloroform extract of IISR Malabar Excel, P. longum, P. chaba, P. colubrinum and hexane extract of P. colubrinum were screened as potent cytotoxic extracts. All the potential extracts, along with synthetic anticancer drug Doxorubicin were again put for MTT assay with a mass range of 5- 100 µg for three time intervals (24, 48 and 72 hrs) and their  $IC_{50}$  was calculated and compared (Table 4 B). Results showed more cytotoxicity with more extract and increased time of exposure with CaSki. Chloroform extract of P. longum and P. colubrinum were found to be highly toxic to CaSki for all the three time intervals. Chloroform extract of IISR Malabar Excel was also toxic to CaSki for 24, 48 and 72 hrs. Hexane extract of P. colubrinum had almost similar toxicity to CaSki as that of chloroform extract of IISR Malabar Excel. P. chaba was less toxic in 24 and 48 hrs but highly toxic in 72 hrs.  $IC_{50}$  for Doxorubicin was  $<5\mu g$ for all the three time intervals. The  $IC_{50}$  value for certain extracts and Doxorubicin were beyond the adopted mass range. So, further experiment has to be performed to find out their exact  $IC_{50}$  values.

*In vitro* cytotoxicity was studied in black pepper extracts for esophageal squamous cell line TE-13 (Dwivedi *et al.*, 2011), breast cancer cell lines like MCF-7, MDA-MB-231 and MDA-MB-468 (Sriwiriyajan *et al.*, 2014), in *P. longum* extracts for lung epithelial adenocarcinoma cell line HCC-827 (Sawhney *et al.*, 2011) and leukaemic cell line K562 (Joy *et al.*, 2010) and in *P. chaba* extracts for large lung carcinoma cell line COR-L23, cervical cancer cell line HeLa and liver cancer cell line HepG2 (Ruangnoo *et al.*, 2012). However, cervical cancer cell line CaSki was least studied in these *Piper* species.

# Preliminary phytochemical analysis of screened extracts

In the case of in vitro antioxidant activity, methanol extract of IISR Malabar Excel and chloroform extract of P. colubrinum showed highest activity. The methanol extract of Panchami also showed high antioxidant activity. In the case of *in vitro* cytotoxicity, chloroform extract of P. colubrinum, IISR Malabar Excel, P. longum, P. chaba and hexane extract of P. colubrinum showed high activities. These screened extracts were tested preliminary for the presence of constituents which may contribute to their high activities. These extracts were tested for alkaloids, flavones, flavonols, phenolics, steroids/triterpenes, saponins, fixed oils and fats, carbohydrates and protein and results are shown in table 5. Alkaloids, phenolics and steroids/triterpenes were present in all the extracts whereas flavonols were present in all the methanol extracts and chloroform extract of P. colubrinum. Flavones were present in all samples except hexane extract of P. colubrinum whereas saponins were present only in methanolic extracts.

Table 4. A:  $IC_{50}$  value for chloroform extract of black pepper varieties and wild *P. nigrum*; B:  $IC_{50}$  value for potential extracts and Doxorubicin

A: IC <sub>50</sub> ( $\mu$ g) for chloroform extract of black pepper varieties and wild <i>P. nigrum</i>						
Sample	24 hrs	48 hrs	72 hrs			
Sreekara	64±2.56°	58±2.32ª	28±1.12 <sup>c</sup>			
Subhakara	64±2.56°	60±2.40 <sup>a</sup>	28±1.12 <sup>c</sup>			
IISR Malabar Excel	60±2.40 <sup>c</sup>	30±1.20 <sup>c</sup>	25±1.00 <sup>d</sup>			
Panchami	72±2.88 <sup>b</sup>	32±1.28°	26±1.04 <sup>cd</sup>			
Panniyur-1	84±3.36ª	52±2.08 <sup>b</sup>	32±1.28 <sup>b</sup>			
IISR Thevam	60±2.40 <sup>c</sup>	32±1.28°	26±1.04 <sup>cd</sup>			
P. nigrum (wild)	86±3.44ª	58±2.32ª	36±1.44ª			
B: $IC_{50}$ (µg) for potential extracts and Doxorubicin						
Sample	24 hrs	48 hrs	72 hrs			
IISR Malabar Excel (Chloroform extract)	60.00±2.4	30.00±1.2	25.00±1.0			
P. chaba (Chloroform extract)	>100	100.0 ±4.0	<5			
P. longum (Chloroform extract)	12.00±0.5	5.2.00±0.2	<5			
P. colubrinum (Chloroform extract)	12.00±0.5	<5	<5			
P. colubrinum (Hexane extract)	56.00±2.2	32.00±1.3	25.00±1.0			
Doxorubicin	<5	<5	<5			

Table 5. Preliminary phytochemical analysis of screened extracts

Phytochemicals	Chemical Test	Screened extracts						
Tested		MM	MP	СС	СМ	CL	СН	HC
Alkaloids	Meyer's Test	+	+	+	+	+	+	+
Flavones	Test with NaOH	+	+	+	+	+	+	-
Flavonols	Shinoda's Test	+	+	+	-	-	-	-
Phenolics	Ferric chloride Test	+	+	+	+	+	+	+
Steroids/triterpenes	Salkowski Test &	+	+	+	+	+	+	+
	Liebermann-Burchard Test							
Saponins	Foam Test	+	+	-	-	-	-	-
Fixed oils and fats	Saponification Test	-	-	-	-	-	-	+
Carbohydrates	Molisch's Test	+	+	-	-	+	+	-
Protein	Biuret Test	-	-	-	-	-	-	-

\*MM- Methanol extract of IISR Malabar Excel, MP-Methanol extract of Panchami, CC-Chloroform extract of *P. colubrinum*, CM-Chloroform extract of IISR Malabar Excel, CL- Chloroform extract of *P. longum*, CH-Chloroform extract of *P. chaba*, HC-Hexane extract of *P. colubrinum*.

\*\* (+) for presence, (-) for absence.

Fixed oils and fats were present only in hexane extract of *P. colubrinum*. Carbohydrates were present in all methanol extracts and chloroform extract of *P. longum* and *P. chaba*. None of the sample has given positive result for protein. Based on these qualitative analysis it was found that potent extracts are good source for various phytochemicals.

#### Conclusion

In vitro antioxidant activity and cytotoxicity of sequential extracts of four important *Piper* species viz. *P. nigrum, P. chaba, P. longum* and *P. colubrinum* and six black pepper varieties viz. Sreekara, Subhakara, IISR Malabar Excel, Panniyur-1, Panchami and IISR Thevam were performed. It can be concluded that methanol and chloroform extracts predominated for antioxidant activity. Considering the four *in*  vitro antioxidant systems tested, methanol extract of IISR Malabar Excel and chloroform extract of P. colubrinum were screened as extracts with highest antioxidant activity. Significant positive correlation was obtained for total phenolic content of methanol and chloroform extracts with their antioxidant activity by all the four assays. In vitro cytotoxicity of the extracts on cervical cancer cell line CaSki showed that chloroform extracts of all the samples and hexane extract of P. colubrinum showed more cytotoxicity. Among the potential extracts, chloroform extracts of *P. longum* and *P. colubrinum* were highly toxic to CaSki. By considering both in vitro antioxidant activity and cytotoxicity, chloroform extract of P. colubrinum was found to be more active than other extracts. Qualitative analysis showed that chloroform extracts of P. colubrinum is a good source of high value constituents. Hence further studies can be

performed for identification of active phytochemical constituents from this extract. This is the first report regarding *in vitro* antioxidant activity and cytotoxicity on CaSki for sequential extracts of *P. colubrinum* fruits cultivated in India. This is also the first report on variability in antioxidant activity and cytotoxicity (on CaSki) of sequential extracts from black pepper varieties selected for the study.

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#### References

- Agbor, A.G., Vinson, A.J., Oben, E.J. and Ngogang, Y.J. 2006. Comparative analysis of the *in vitro* antioxidant activity of white and black pepper. Nutrition Research 26(12): 659-663.
- Anshuman, P.S., Sing, K.P. and Asra, K.G. 1984. Effect of Vardhaman Pippali (*Piper longum*) on patients with respiratory disorders. Sachitra Ayurved 37(1): 47-49.
- Asimi, O.A., Sahu, N.P. and Pal, A.K. 2013. Antioxidant capacity of crude water and ethyl acetate extracts of some Indian spices and their antimicrobial activity against *Vibrio vulnificus* and *Micrococcus luteus*. Journal of Medicinal Plants Research 7(26):1907-1915.
- Bano, G., Raina, R.K., Zutshi, U., Bedi, K.L., Johri, R.K. and Sharma, S.C.1991.Effect of piperine on bioavailability and pharmacokinetics of propranolol and theophylline in healthy volunteers. European Journal of Clinical Pharmacology 41(6): 615-617.
- Barua, C.C., Singh, A., Sen, S., Barua, A.G. and Barua, I.G. 2014. *In vitro* antioxidant and antimicrobial activity of seeds of *P.longum* Linn: A comparative study. SAJ Pharmacy and Pharmacology 1(1): 1-10.
- Biswas, S.M., Chakraborty, N., Chakraborty, P. and Sarkar, S. 2012. Antioxidant and antimicrobial activities of hot pungent chabbarin are responsible for the medicinal properties of Piper chaba Hunter. Research Journal of Medicinal Plant 6(8): 574-586.
- Braca, A., Tommasi, N.D., Bari, L.D., Pizza, C., Politi, M. and Morelli, I. 2001. Antioxidant principles from Bauhinia terapotensis. Journal of Natural Products 64: 892-895.
- Carter, P. 1971. Spectrophotometric determination of serum iron at the submicrogram level with a new reagent (Ferrozine). Analytical Biochemistry 40: 450-458.
- Chatterjee, S., Niaz, Z., Gautam, S., Adhikari, S., Variyar, P.S. and Sharma, A. 2007. Antioxidant activity of some phenolic constituents from green pepper (*Piper nigrum* L.) and fresh nutmeg mace (*Myristica* fragrans). Food Chemistry 101: 515-523.

- Chen, X. and Ahn, D.U. 1998. Antioxidant activities of six natural phenolics against lipid oxidation induced by Fe<sup>2+</sup> or ultra violet light. Journal of American Oil Chemists' Society 75: 1717-1721.
- Demiray, S., Pintado, M.E. and Castro, P.M.L. 2009. Evaluation of phenolic profiles and antioxidant activities of Turkish Medicinal Plants: *Tilia argentea*, *Crataegi folium* leaves and *Polygonum bistorta* roots. World Academy of Science, Engineering and Technology 54: 312-317.
- Dwivedi, V., Shrivastava, R., Hussain, S., Ganguly, C. and Bharadwaj M. 2011. Cytotoxic potential of Indian spices (extracts) against esophageal squamous carcinoma cells. Asian Pacific journal of cancer prevention 12: 2069-2073.
- Freshney, I.R. 2007. Culture of animal cells: A manual of basic technique and specialized applications. 6th ed. USA: John Wiley & Sons, Inc.
- Gopalakrishna, A.G, Lokesh, B.R, Sugasini, D. and Kancheva, V.D. 2010. Evaluation of the antiradical and antioxidant properties of extracts from Indian red chili and black pepper by *in vitro* models. Bulgarian Chemical Communications 42(2): 62-69.
- Gulcin, I. 2005. The antioxidant and radical scavenging activities of black pepper (*P. nigrum*) seeds. International Journal of Food Sciences and Nutrition 56 (7): 491-499.
- Gupta, K.R., Chawla, P., Tripathi, M., Shukla, K.A. and Pandey, A. 2014. Synergistic antioxidant activity of tea with ginger, black pepper and tulsi. International Journal of Pharmacy and Pharmaceutical Sciences 6 (5): 477-479.
- Joy, B., Sandhya, C.P. and Remitha, K.P. 2010. Comparison and bioevaluation of *P. longum* fruit extracts. Journal of Chemical and Pharmaceutical Research 2(4): 612-622.
- Kapoor, I.P.S., Singh, B., Singh, G., De Heluani, C.S., De Lampasona, M.P. and Catalan, C.A.N. 2009. Chemistry and *in vitro* antioxidant activity of volatile oil and oleoresins of black pepper (*Piper nigrum*). Journal of Agricultural and Food Chemistry 57: 5358-5364.
- Karsha, P.V. and Lakshmi, O.B. 2010. Antibacterial activity of black pepper (*Piper nigrum* Linn.) with special reference to its mode of action on bacteria. Indian Journal of Natural Products and Resources 1(2): 213-215.
- Khandelwal, K.R. 2008. Practical Pharmacognosy: Techniques and experiments. 19<sup>th</sup> ed. Pune: Nirali Prakashan.
- Kokate, C.K., Purohit, A.P. and Gokhale, S.B. 2008. Pharmacognosy, 28<sup>th</sup> ed. Pune: Nirali Prakashan.
- Krishnaswami, T., Sellamuthu, M. and Subramaniam, P. 2013. *In vitro* radical scavenging potential of pod and seed extracts of *Bauhinia tomentosa* L. International Journal of Pharmacy and Pharmaceutical Sciences 5 (1): 346-351.
- Lee, E.B., Shin, K.H. and Woo, W.S. 1984. Pharmacological study of piperine. Archives of Pharmacal Research 7: 127-132.
- Luo, Q., Sun, M. and Corke, H. 2004. Antioxidant

activity and phenolic compound of 112 traditional Chinese Medicinal plants associated with anticancer. LifeSciences 74: 2157-2184.

- Maizura, M., Aminah, A. and Wan Aida, W.M. 2011. Total phenolic content and antioxidant activity of Kesum (*Polygonum minus*), ginger (*Zingiber officinale*) and turmeric (*Curcuma longa*) extract. International Food Research Journal 18: 529-534.
- Malick, C.P. and Singh, M.B. 1980. Plant enzymology and histoenzymology. New Delhi: Kalyani Publishers.
- Mariutti, L.R.B., Barreto, G.P.M., Bragagnolo, N. and Mercadante, A.Z. 2008. Free radical scavenging activity of ethanolic extracts from herbs and spices commercialized in Brazil. Brazilian Archives of Biology and Technology 51(6):1225-1232.
- Mosmann, T. 1983. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. Journal of Immunological Methods 65(1-2): 55-63.
- Nahak, G. and Sahu, R.K. 2011. Phytochemical evaluation and antioxidant activity of *Piper cubeba* and *Piper nigrum*. Journal of Applied Pharmaceutical Science 01(08): 153-157.
- Nooman, A.K., Ashok, K.S., Atif, A.O., Zaha, E.A. and Husni, F. 2008. Antioxidant activity of some common plants. Turkish Journal of Biology 32: 51-55.
- Oyaizu, M. 1986. Studies on products of browning reaction: antioxidative activities of products of browning reaction prepared from glucosamine. Japanese Journal of Nutrition and Dietetics 44 (6):307-15.
- Pietta, P.G. 2000. Flavonoids as antioxidants. Journal of Natural Products 63: 1035-1042.
- Pingili, D., Anarthe, J.S. and Raghavendra, M.N. 2012. Evaluation of the polyherbal extract for antioxidant, anticancer and antidiabetic activity. Annals of Phytomedicine 1(1): 39-45.
- Policegoudra, R.S., Chandrashekhar, R.H., Aradhya, S.M. and Singh, L. 2011. Cytotoxicity, platelet aggregation inhibitory and antioxidant activity of *Curcuma amada* Roxb. extracts. Food Technology and Biotechnology 49(2):162–168.
- Prasad, M.P. and Sushant, S. 2014. *In vitro* antioxidant potential analysis of some medicinal plant species found in southern India. Indian Journal of Advances in Plant Research 1(2): 27-30.
- Prieto, P., Pineda, M. and Aguilar, M. 1999. Spectrophotometric quantitation of antioxidant capacity through the formation of phosphomolybdenum complex: Specific application to the determination of vitamine E. Analytical Biochemistry 269: 337-341.
- Rahman, A., Al-Reza, S.M., Sattar, M.A. and Kang, S.C. 2011. Potential roles of essential oil and extracts of *Piper chaba* Hunter to inhibit Listeria monocytogenes. Records of Natural Products 5(3): 228-237.
- Ramesh kumar, K.B., Anu aravind, A.P. and Mathew, P.J. 2011. Comparative phytochemical evaluation and antioxidant assay of *Piper longum* L. and *Piper chaba* Hunter used in Indian traditional systems of medicine. Journal of Herbs, Spices & Medicinal Plants 17: 351-360.

- Ravindran, P.N. and Remashree, A.B. 1998. Anatomy of Piper colubrinum Link. Journal of Spices and Aromatic Crops 7(2): 111-123.
- Rice-Evans, C.A., Miller, J. and Paganga, G. 1997. Antioxidant properties of phenolic compounds. Trends in Plant Science 2:152-159.
- Ruangnoo, S., Itharat, A., Sakpakdeejaroen, I., Rattarom, R., Tappayutpijam, P. and Pawa, K.K. 2012. *In vitro* cytotoxic activity of Benjakul herbal preparation and its active compounds against human lung, cervical and liver cancer cells. Journal of the Medical Association of Thailand 95(1): 127-134.
- Samudram, P., Vasuki, R., Rajeshwari, H., Geetha, A. and Moorthi, P.S. 2009. Antioxidant and antihepatotoxic activities of ethanolic crude extract of *Melia* azedarach and *Piper longum*. Journal of Medicinal Plants Research 3(12): 1078-1083.
- Santoso, J., Yoshie-Stark, Y. and Suzuki, T. 2004. Antioxidant activity of methanol extracts from Indonesian seaweeds in an oil emulsion model. Fisheries Science 70(1):183-188.
- Sawhney, S.S., Painuli, R.M. and Chauhan, N. 2011. Evaluation of bactericidal and anticancer properties of fruits of *Piper longum*. International Journal of Pharmacy and Pharmaceutical Sciences 3(5):282-287.
- Shan, B., Cai, Y.Z., Sun, M. and Corke, H. 2005. Antioxidant capacity of 26 spice extracts and characterization of their phenolic constituents. Journal of Agricultural and Food Chemistry 53: 7749-7759.
- Singh, R., Singh, N., Saini, B.S. and Rao, S.H. 2008 *In vitro* antioxidant activity of pet ether extract of black pepper. Indian Journal of Pharmacology 40(4): 147-151.
- Srinivasa, P. R., Jamil, K., Madhusudhan, P., Anjali, G. and Das, B. 2001. Antimicrobial activity of isolates from *Piper longum* and *Taxus baccata*. Pharmaceutical Biology 39(3): 236-238.
- Sriwiriyajan, S., Ninpesh, T., Sukpondma, Y., Nasomyon, T. and Graidist, P. 2014. Cytotoxicity screening of plants of Genus *Piper* in Breast Cancer Cell Lines. Tropical Journal of Pharmaceutical Research 13(6): 921-928.
- Sunila, E.S. and Kuttan, G. 2004. Immunomodulatory and anti-tumour activities of *Piper longum* Linn. and piperine. Journal of Ethnopharmacology 90(2-3): 339-346.
- Taufiq-Ur-Rahman, Md., Shilpi, J.A., Ahmed, M., Hossain, C.F., 2005. Preliminary Pharmacological studies on Piper chaba stem bark. Journal of Ethnopharmacology 99: 203-209.
- Trease, G.E. and Evans, W.C. 2002. Pharmacognosy. 15<sup>th</sup> ed. London: Saunders Publishers.
- Van Acker, S.A.B.E., Van Balen, G.P., Van Den, B.D.J., Bast, A. and Van Der Vijgh, W.J.F.1998. Influence of iron chelation on the antioxidant activity of flavonoids. Biochemical Pharmacology 56: 935-943.
- Van Acker, S.A.B.E., Van Den, B.D.J., Tromp, M.N.J.L., Griffioen, D.H., Van Bennekom, W.P., Van Der Vijgh, W.J.F. and Bast, A. 1996. Structural aspects of antioxidant activity of flavonoids. Free Radical

Biology and Medicine 20: 331-342.

- Vijayakumar, R.S., Surya, D. and Nalini, N. 2004. Antioxidant efficacy of black pepper (*Piper nigrum* L.) and piperine in rats with high fat diet induced oxidative stress. Redox Report 9(2): 105-110.
- Williams, R.J., Spencer, J.P. and Rice-Evans, C. 2004. Flavonoids: antioxidants or signaling molecules. Free Radical Biology and Medicine 36: 838-849.
- Wu, C.Q., Chen, F., Wang, X., Kim, H.J., He, G.Q., Haley-Zitlin, V. and Huang, G. 2006. Antioxidant constituents in feverfew (Tenacetum parthenium) extract and their chromatographic quantification. Food Chemistry 96: 220-227.
- Yanishlieva, N.V. and Marinova, E.M. 2001. Stabilization of edible oils with natural antioxidants. European Journal of Lipid Science and Technology 103: 752-767.
- Yusuf, M., Chowdhury, J.U., Wahab, M.A. and Begum, J. 1994. Medicinal plants of Bangladesh. Dhaka, Bangladesh: BCSIR.